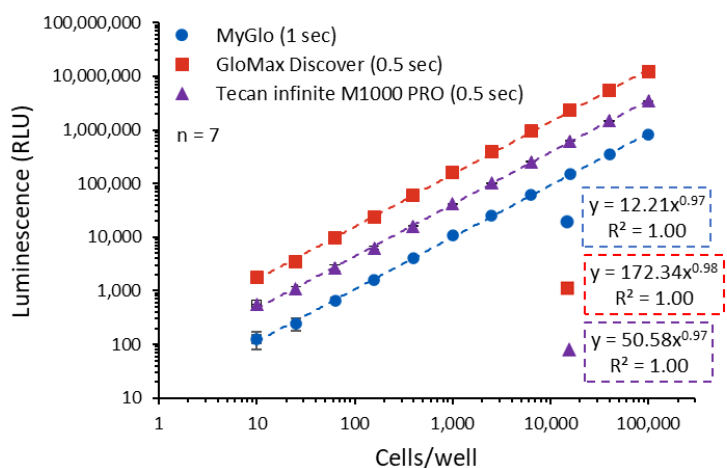
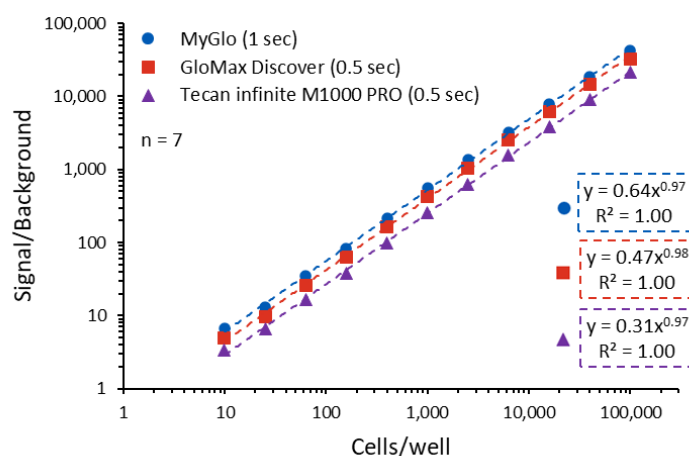


Comparison among MyGlo, GloMax Discover and Tecan Infinite

CellTiter-Glo vs Jurkat cell number
(all white 96-well plate)

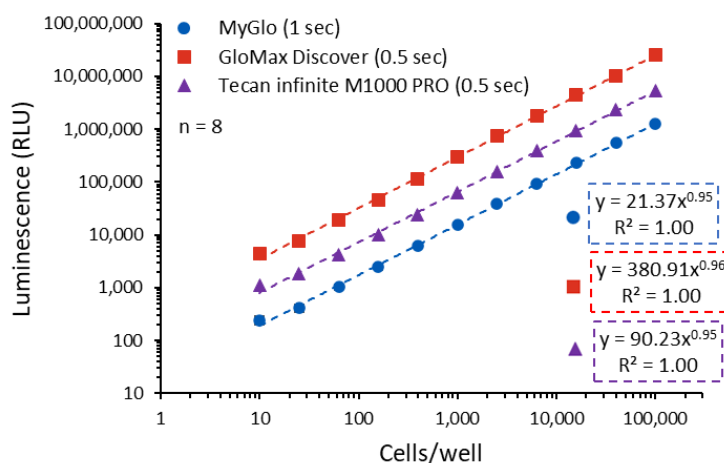


CellTiter-Glo vs Jurkat cell number
(all white 96-well plate)

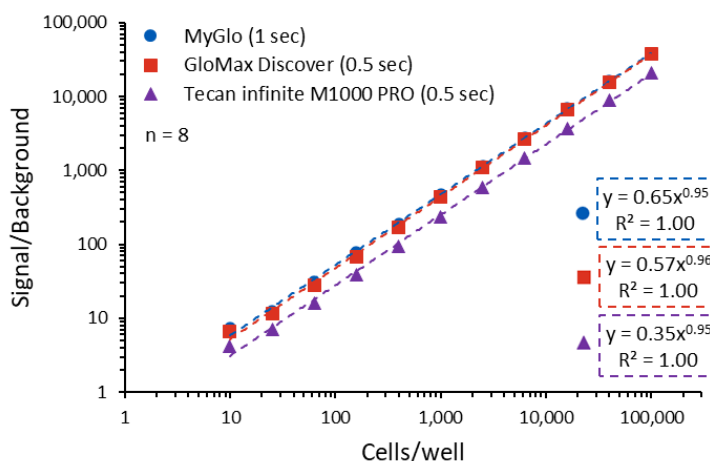


Using CellTiter®-Glo Luminescent Cell Viability Assay

CellTiter-Glo 2.0 vs Jurkat cell number
(all white 96-well plate)



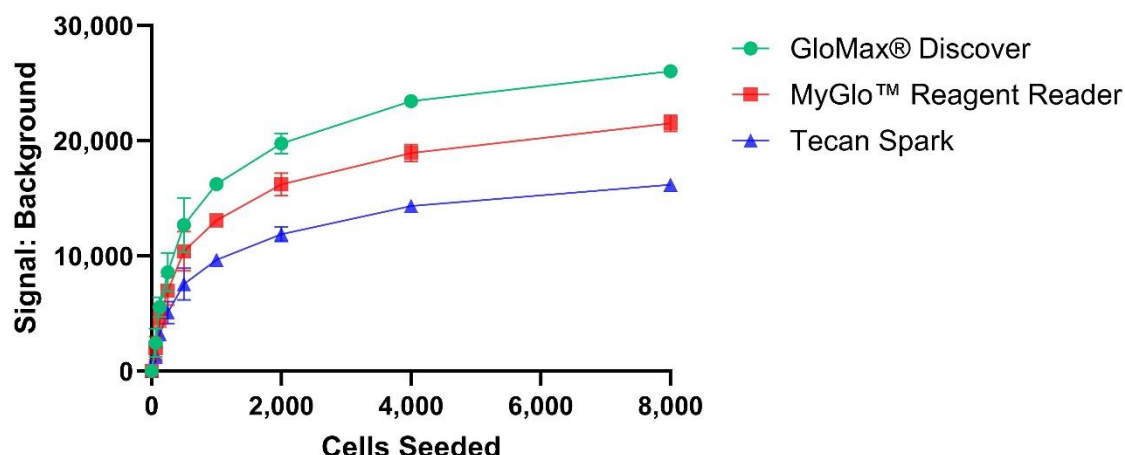
CellTiter-Glo 2.0 vs Jurkat cell number
(all white 96-well plate)



Using CellTiter®-Glo 2.0 Cell Viability Assay

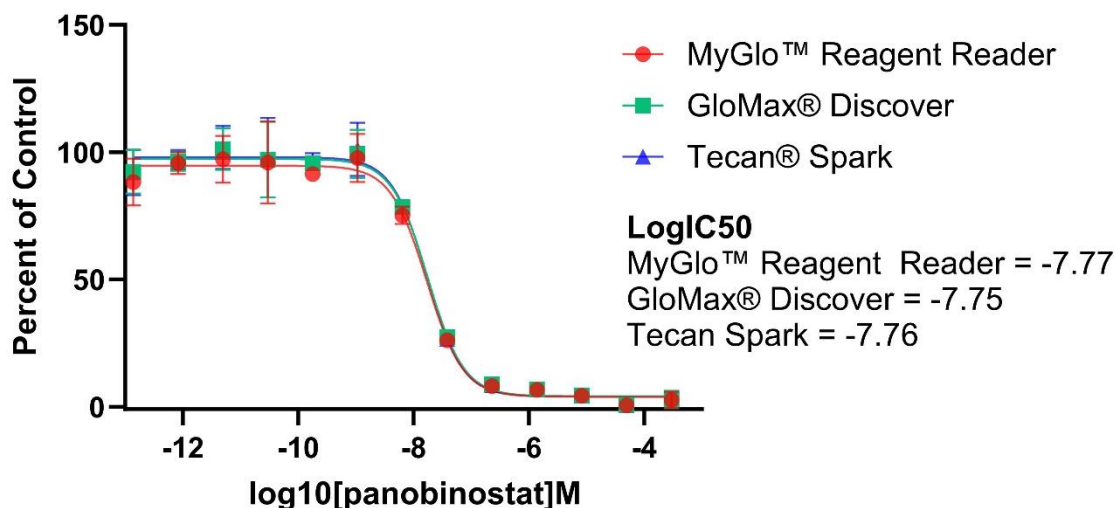
Comparison among MyGlo, GloMax Discover and Tecan Spark

Cell-Titer-Glo® 3D with HCT116 spheroids



Signal to background comparison between three instruments using CellTiter®-Glo 3D. Signal:background luminescence data from CellTiter®-Glo 3D with a titration of HCT116 cells seeded from 8,000 cells/well to 125 cells/well in ULA, round-bottom 96-well plates, then grown 3.5 days to form spheroids. The same plate was read on the GloMax® Discover Microplate Reader (green circles), MyGlo™ Reagent Reader (red squares) and Tecan Spark® (blue triangles). Points represent the mean and the error bars represent the standard deviation of six replicates.

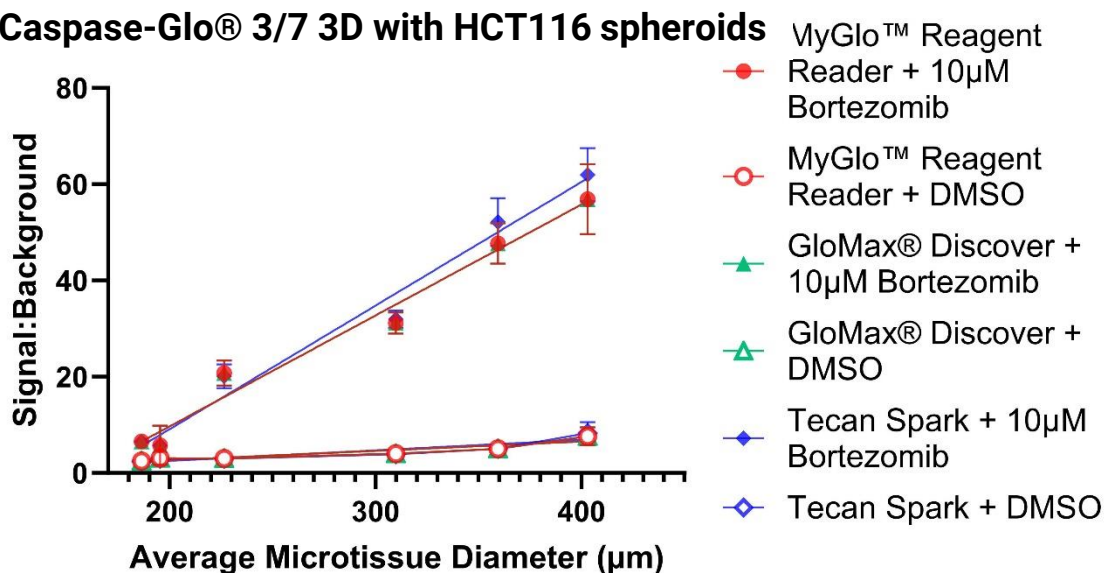
HCT116 spheroids dosed for 48 hours



Dose response curve from GraphPad Prism of CellTiter®-Glo 3D with HCT116 spheroids treated with Panobinostat for 48 hours. 200 HCT116 cells were seeded, grown for 3.5 days in round-bottom ULA plates to generate spheroids (average 230µm diameter) and treated with varying concentrations of Panobinostat for 48 hours. Percent viability (% of control) is calculated by taking each compound concentration replicate Net RLU and dividing by the Net RLU average of control wells (DMSO treatment), then multiplied by 100. This is the percent of control per sample replicate. The percent of control of the same compound replicates is then averaged. The mean percent of control is shown and the error bars represent the standard deviation of 3 replicates. Output in GraphPad Prism from the GloMax® Discover Microplate Reader (green squares), MyGlo™ Reagent Reader (red circles) and Tecan Spark® (blue triangles). LogIC50 values are included on the graph.

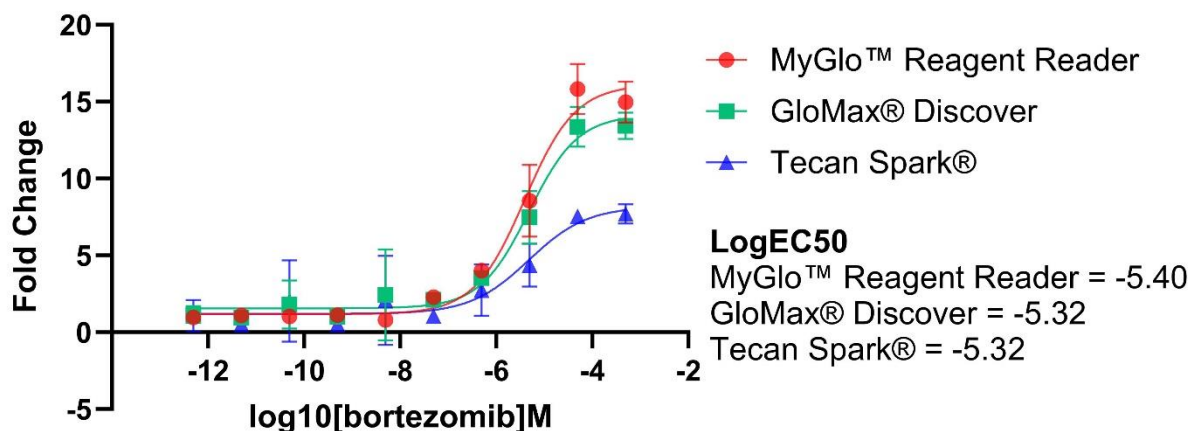
Comparison among MyGlo, GloMax Discover and Tecan Spark

Caspase-Glo® 3/7 3D with HCT116 spheroids



Signal:background comparison between three instruments using Caspase-Glo® 3/7 3D. Signal:background luminescence data from Caspase-Glo® 3/7 3D with HCT116 spheroids generated over 3.5 days in ULA plates from cells seeded at 1,000 cells/well (producing ~403μm diameter spheroids) to 32.5 cells/well (producing ~186μm diameter spheroids), then treated for 24h with 10μM Bortezomib to induce apoptosis or DMSO as an uninduced control. The same plate was read on the GloMax® Discover Microplate Reader (green triangles), MyGlo™ Reagent Reader (red circles) and Tecan Spark® (blue diamonds). Points represent the mean and the error bars represent the standard deviation

HCT116 spheroids dosed for 48 hours



Dose response curve from GraphPad Prism of Caspase-Glo® 3/7 3D with 400 seeded HCT116 cells treated with Bortezomib for 24 hours. 400 HCT116 cells were seeded, grown for 3.5 days in round-bottom ULA plates to generate spheroids (475μm diameter) and then treated with varying concentrations of Bortezomib for 24 hours. Fold change is the sample replicate RLU average (background subtracted) divided by the control (DMSO) replicate RLU average (also background subtracted). The mean fold change is shown and the error bars represent the standard deviation of n=3. Output in GraphPad Prism from the GloMax® Discover Microplate Reader (green squares), MyGlo™ Reagent Reader (red circles) and Tecan Spark® (blue triangles). LogEC50 values are included on the graph.